

International Journal of Pharmaceutical and Medicinal Research

Journal homepage: www.ijpmr.org

Review Article

Pharmacological activities of Abrus precatorius (L.) seeds

Meena Prabha. P^{*1}, Chendraya Perumal. P², Praveen Kumar. M³, Soundarrajan⁴. S, Srinivasan. M⁵, R. Sampathkumar⁶

^{1,2,3,4,5}Department of Pharmacognosy, J.K.K. Nattraja College of Pharmacy, Kumarapalayan-638183, Namakkal (Dt), Tamilnadu. ⁶Principal & Head Department of Pharmaceutics, J.K.K. Nattraja College of Pharmacy, Kumarapalayan 638183, Namakkal (Dt), Tamilnadu

ARTICLE INFO:	ABSTRACT
Article history: Received: 08 April, 2015 Received in revised form: 24 April, 2015 Accepted: 28 April, 2015 Available online: 30 April, 2015	<i>Abrus precatorius</i> (L) is herbal plant belonging to the family of fabaceae (Leguminosae)- pea family. This plant is commonly known as Indian liquorices. This plant species has been found to display a wide variety of biochemical activities. All parts of the plants have medicinal properties so it is a very valuable medicinal plant which is utilized in traditional system of medicine. Seeds are used as a poultice in the vagina in Ayurvedic and Unani medicine as an abortifacient and it also reported to posses antidiarrheal, antifertility, antispasmodic, antiyeast, antidiabetic, embryo toxic, mitogenic activity, protease(HIV) inhibition, antigonadotropin, agglutinin activity, antibacterial, antioxidant, anticataractic and teratogenic effect. This crab's eye view mainly on the pharmacogostic characteristics and pharmacological actions of the plant.
<i>Keywords:</i> <i>Abrus precatorius</i> Fabaceae Abortifacient Mitogenic activity	

1. Introduction

Abrus precatorius (L.) is a popular medicinal plant belonging to the family of fabaceae (Leguminosae)- pea family. Many medicinal uses are ascribed to this plant. The leaves, stem and roots are sweet-tasting due to the presence of glycyrrhizin, of which about 9–10 % is in the leaf and is an ornamental, twining, woody vine which grows to a height of 10 to 20 feet when supported by other plants. Leaves are alternate, compound, feather-like (pinnately divided), with small oblong leaflets. Flowers are numerous and appear in the leaf axils along the stems. They are small and occur in clusters 1 to 3 inches long, usually red to purple, or occasionally white. The fruit is a legume (pea shaped pod) about 3 cm long containing hard ovoid seeds about 1 cm long. This plant is commonly known as Indian liquorices. The most common variety of seed is glossy, bright scarlet, with the area around the hilum (point of attachment) being black. Most cases of poisoning involve the ingestion (inadvertently or deliberately) of these attractive red seeds. However, there are other less common varieties of this plant that produce different coloured seeds: for instance, black with a white spot, and white with a black spot[1].

Abrus precatorius is a plant that originates from Southeast Asia and now can be found in subtropical areas of the world. The name *Abrus*, meaning beautiful or graceful, is used to describe the appearance of the seed. The seed is found in a variety of colors such as black, orange, and most commonly, red with a glossy appearance with the black band at the end that attaches to the plant. The seeds are used in a variety of jewelry, trinkets, and ornaments; the *Abrus* seed itself is known by a variety of names that include rosary pea, prayer bead, and jequirity bean. Precare (from which the species name is derived) meaning to pray, references its common use in rosaries.

The seeds of *Abrus precatorius* have been used through history in a variety of roles. Due to their uniform size and weight, they were once known as rati, and used as weights for weighing gold and silver. Formerly Indians used these seeds to weigh gold using a measure called a Ratti, where 8 Ratti = 1 Masha; 12 Masha = 1 Tola (11.6 Grams). The *Abrus* seeds have also been used for medicinal purposes, including the treatment of chronic eye disease. Arabic culture has purportedly used the seed as an aphrodisiac known as coq's eye. The toxicity of the *Abrus* seed was associated with its use as a fish poison as well as a homicidal agent.

The poisoning by the seeds of *Abrus precatorius* has been reviewed and reported often in literature. Death has been reported with twenty seeds bended with water. The symptoms included vomiting of blood, severe pain in the eyes and burning of ears. Death ensued in two days[2]. Death in children has been reported from ingestion of one or two seeds[3]. There are reports of fatal outcomes of men, who ate one or two beans only.

*Corresponding author: Meena Prabha. P, Department of Pharmacognosy, J.K.K. Nattraja College of Pharmacy, Kumarapalayan-638183, Namakkal (Dt), Tamilnadu., India.; Tel.: 08973750825; E-mail: <u>Meenaprabha.p@jkkn.org</u>

Swallowing of intact beans is nearly harmless. Boiled seeds eaten by the residents of the Andaman Islands were harmless, too. They were analyzed for proteins, amino acid composition, minerals and antinutrional factors with positive results[4]. In the seeds the toxic principle is abrin, a mixture of at least five lectins, abrin A - D, and abrus-agglutinin. The abrins consist of two peptide chains connected by a disulfide bridge. Abrin A consists of an A-chain with N-glycosidase activity, which inhibits protein synthesis, and lectin-like B-chain responsible for binding with cell-surface receptors and penetrating of abrin-A molecule into the cell[5].

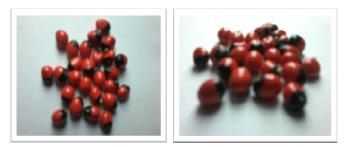


Figure 1: Abrus precatorius (L)

2. Pharmacological activities

Abortifacient effect

Chloroform/methanol extract of seeds, administered subcutaneously to rats at a dose of 50.0 mg/animal, was inactive. Water extract of dried seeds, administered intragastrically to pregnant rats at a dose of 125.0 mg/kg, was active[11]. Ethanol (95%) extract of seeds, administered orally at a dose of 200.0 mg/kg, was inactive on pregnant hamsters and active on pregnant rats[12]. Petroleum ether extract of seeds, administered orally to rats, was inactive[13].

Agglutinin activity

Water extract of fresh seeds, in cell culture at a concentration of 2.0 microliters/ml, was active on human lymphocytes[14].

Alkaline phosphatase inhibition

Petroleum ether extract of seed oil, administered orally, was active on the uterus of rats[15].

Anthelmintic activity

Water extract of dried seeds produced weak activity on *Caenorhabditis elegans*, LC50 15.8 mg/ml[16].

Antidiarrheal activity

Chromatographic fraction of dried seeds, administered intragastrically to rats at a dose of 10.0 mg/kg, was active vs castor oil-induced diarrhea[17].

Antifertility effect

Chloroform/methanol extract of seeds. administered subcutaneously to female rats at a dose of 50.0 mg/ animal, was active[18]. Ethanol (80%) extract of seeds, administered orally and subcutaneously to female rats at a dose of 1.0 mg/animal, was inactive[19]. Ethanol (95%) and water extracts of seeds, administered orally to mice, were inactive, and petroleum ether extract was active[20]. Ethanol (95%), water and petroleum ether extracts of leaves, administered orally to female mice, were extract of seeds, inactive^[20]. Ethanol administered intragastrically to male rats at a dose of 100.0 mg/kg for 60 days, was active. There was a significant decrease in the number of pregnant Female[21]. Ethanol/water (1:1) extract of dried seeds, administered by gastric intubation to male rats at a dose of 250.0 mg/kg, was active. No pregnancies were reported for the 20 females paired with 10 males treated for 60 days; mating probably occurred in all cases, but this is not entirely clear. Pregnancies were again reported after withdrawal of treatment[22]. Hot water extract of dried plant, administered orally to human females at a dose of 0.28 gm/person, was active. The extract was administered as a mixture of Embelia ribes (fruit), Piper longum (fruit), Ferula assafoetida, Piper betele, Polianthes tuberose and Abrus precatorius. One dose was taken, starting from the second day of menstruation, twice daily for 20 days. Sexual intercourse was avoided during the dosing period. The treatment is claimed effective for 4 months. The biological activity has been patented [23]. Seed oil, administered orally to female mice at a dose of 25.0 mg/ animal, to female mice, and to rats at a dose 150.0 mg/animal, was active. No control animal was used[24].

Antigonadotropin effect

Ethanol (95%) extract of dried seeds, administered by gastric intubation to mice at a dose of 150.0 mg/kg, was active[25].

Anti-implantation effect

Chloroform/methanol (2:1) extract of seeds, administered subcutaneously to pregnant rats at a dose of 50.0 mg/animal, was active[18]. Ethanol (95%) extract of seeds, administered orally to rats and hamsters at a dose of 200.0 mg/kg, was inactive[12].

Antispasmodic activity

Chromatographic fraction (a gel filtration fraction from a methanol-water (1:1) extract) of seeds, at a concentration of 0.2 mg/ml, was active on the uterus of rats vs PGE-2-,ACh-, oxytocin- and epinephrine-induced contractions[26].

Antispermatogenic effect

Ethanol extract of seeds, administered intragastrically to male rats at a dose of 100.0 mg/kg for 60 days, was inactive[21]. Ethanol/water (1:1) extract of dried seeds, administered by gastric intubation to rats at a dose of 250.0 mg/ kg, was active. Although no significant histologic changes in the testes were reported, sperm concentration was reported to be significantly decreased in both cauda epididymis and testes after dosing for

60 days[22]. Sterol fraction of dried seeds administered intramuscularly to rats was active. Testicular lesions marked by the cessation of spermatogenesis and a significant reduction in the diameter of the seminiferous tubules were also noted[23].

Antiyeast activity

Dried seeds at a concentration of 1.0% on agar plate were active on *Cryptococcus neoformans*[28].

Contraceptive and/or interceptive effect

Petroleum ether extract of seed oil, administered orally to rats, was active[15].

Embryotoxic effect

Ethanol (95%) extract of seeds, administered orally to pregnant hamsters and rats at doses of 200.0 mg/kg, was inactive[12]. Petroleum ether extract, administered orally to rats at a dose of 150.0 mg/kg, was inactive[13]. Water extract of dried seeds, administered intragastrically to pregnant rats at a dose of 125.0 mg/kg, was inactive[30].

Estrous cycle disruption effect

Seeds, administered orally to female rats at doses of 0.05, 0.5, and 5.0 mg/animal, were inactive[**31**]. Chloroform/methanol (2:1) extract of seeds, administered subcutaneously to rats at a dose of 1.0 mg/animal, was active[**19**]. Seeds, administered by gastric intubation to rats at doses of 10.0, 5.0, and 2.0 gm/kg, were active; 80, 50, and 25%, respectively, of the rats depicted extensive leukocytic smears, but with no significant effect on uterine weight[**32**].

Hemagglutinin activity

Water extract of seeds was active on the red blood cells of ant (leafcutter), buffalo, cat, chicken, dog, duckling, guinea pig, horse, human adult (blood groups A, B, and O), lamb, mice, pigeon, rabbit, rat, and ox; weakly active on cow and ewe and inactive on goat[23,24].

Insect sterility induction

Petroleum ether extract of dried seeds, applied externally at a concentration of 1.0 microliter, was active on *Dysdercus cingulatus*. The extract was active in males alone. The saline extract produced weak activity in both males and females[35].

Intestinal fluid retention effect

Chromatographic fraction of dried seeds, administered intragastrically to rats at a dose of 10.0 mg/kg, was active on the small intestine vs PGE2-induced enteropooling. Effect assayed 30 minutes after oral dose of PGE2[17].

Chromatographic fraction of dried seeds, administered intragastrically to rats at a dose of 10.0 mg/ kg, was active. Effect was not as great as that of an equal amount of atropine[17].

Luteal suppressant effect

Chloroform/ methanol (2:1) extract of seeds, administered subcutaneously to rats at a dose of 50.0 mg/animal, was active[18].

Mitogenic activity

Water extract of fresh seeds, in cell culture at a concentration of 2.0 microliters/ml, was inactive on human lymphocytes[36].

Protease (HIV) inhibition

Water and methanol extracts of dried seeds were inactive, IC50 > 500 mcg/ml[37].

Reverse transcriptase inhibition

Water and methanol extracts of commercial sample of seeds, in cell culture, were inactive on virus-avian myeloblastosis, IC50 > 1000 mg/ml[38].

Smooth muscle stimulant activity

Chromatographic fraction (gel filtration 4–9 of a methanol-water (1:1) extract of seeds, at a concentration of 0.2 mg/ml, was active on guinea pig ileum; a concentration of 0.5 mg/ ml, was active on the stomach of rats.[26] Seed oil, at a concentration of 1.8 mcg/ml, was active on the ileum of guinea pigs[**39**].

Spermicidal effect

Ethanol extract of seeds, administered intragastrically to male rats at a dose of 100.0 mg/kg for 60 days, was active. Impaired sperm motility and structural abnormalities of sperm were observed. Sperm ATPase level was decreased[21]. Ethanol/water (1:1) extract of dried seeds was active on the sperm of rats. There was a decrease in motility when sperm was mixed with the extract. When administered by gastric intubation, at a dose of 250.0 mg/kg, there was a large decrease in motility of sperm from the cauda epididymis of the rats given the extract for 60 days[22].

Teratogenic activity

Water extract of dried seeds, administered intragastrically to pregnant rats at a dose of 125.0 mg/kg, was active[**30,11**].

Toxic effect (general)

Seeds, administered orally to horses at a dose of 15.0 gm, were active. Tolerance developed when small, incrementally-

increased doses were given [40]. Seeds, at a concentration of 0.5% of diet in chicken, were active. Chickens were fed a mixture of Abrus precatorius seeds and Cassia senna fruit. Toxicity included catarrhal enteritis, hepatocellular necrosis, reduced weight, and anemia[41]. Ethanol (95%) extract of seeds, administered subcutaneously to male mice at a dose of 500.0 mg/kg, was active. One hundred percent mortality was observed within 48-49 hours AP028. Seeds, administered orally to human adults, were active. Severe gastroenteritis, multiple serosal hemorrhages, swelling and inflammation of the Peyer's patches, swelling and inflammation of retroperitoneal lymph nodes, focal necrosis in the liver and kidneys, retinal hemorrhages early in course of intoxication, nausea, vomiting, diarrhea, dehydration, convulsions, and collapse are possible symptoms. Symptoms may begin after delay of up to several days and may persist for as long as 10-11 days. Death in children has been reported from eating 1 or more seeds AP020. Two children who chewed seeds became irrational, had tetany, flushing of skin, widely dilated pupils, and appeared to hallucinate. Treatment with neostigmine and barbiturates was successfulAP042. Seeds, administered subcutaneously to male mice at a dose of 0.90 gm/kg, were active. Forty-four deaths were observed in 5-21 hours[42]. Seeds administered orally to cows at a dose of 0.09 gm/kg were active. Death was observed in 1 of 44 animals. Methanol (75%) extract of dried leaves, administered intragastrically to mice at a dose of 2.0 gm/kg, was inactive[45]. Leaf and stem, administered orally to cows at a dose of 15.4 gm/kg, was inactive[46]. Seeds, in the ration of livestock, were active; nitrate poisoning was observed[47]. Beans, ingested by human adult, produced pulmonary edema and hypertension[48].

Toxicity

Fatal incidents have been reported following ingestion of wellchewed seeds of Abrus precatorius. Because of its hard seed coat, it can pass through the gastrointestinal tract undigested and remain harmless. The unripe seed has a soft and easily broken seed coat and is thus more dangerous. It has been reported that poisoning has been experienced through a finger prick when stringing the seed. Symptoms may develop after a few hours to several days after ingestion. They include severe gastroenteritis with pronounced nausea and vomiting. Mydriasis will occur, as well, as muscular weakness, tachycardia, cold sweat, and trembling. There is no known physiological antidote. The treatment is essentially symptomatic. Since there is a long latent period associated with abrin poisoning, little value can be placed on induction of emesis or gastric lavage; these measures are useful only if ingestion has just occurred. Bismuth trisilicate may be given during poisoning by Abrus precatorius to reduce the degree of gastrointestinal damage. If the emesis and/or diarrhea become excessive, replacement fluids and electrolytes are advocated. If hemorrhage occurs, blood transfusion may be necessary.

Uterine relaxation effect

Chromatographic fraction (a gel filtration fraction from a methanol/water [1:1] fraction) of seeds, at a concentration of 1.1 mg/ml, was active on the uterus of rats[26].

Uterine stimulant effect

Chromatographic fraction (gel filtration fractions 4–9 of a methanol/water [1:1] extract) of seeds, at a concentration of 0.2 mg/ml, was active on the uteri of pregnant and no pregnant rats[26]. Ethanol (95%) extract of dried seed oil, administered intravenously to guinea pigs at a dose of 1000 mcg/ml, produced weak activity.[49] Seed oil, at a concentration of 3.6 mg, was active on the uteri of guinea pigs and rats. The action was blocked by indomethacin but not by atropine AP113. Water extract of seeds was active on the uterus of guinea pig[50].

Antidiabetic effect

Chloroform- methanol extract of seeds of Abrus precatorius produce antidiabetic effect in alloxan induced diabetic in rabbits. Blood glucose level was determined by o-toluidine method[51].

Anti microbial activity

An antimicrobial activity of *Abrus precatorius* seed extract was assayed by in vitro studies in agar well diffusion method against ten bacterial species. Methanol extract exhibited antibacterial activity towards almost all the bacterial microorganisms[52].

Anthelminitic activity

Ethanolic extract of seeds of *Abrus precatorius* shows good anthelminitic activity. Earth warm used for determining activity of anthelminitic activity[53].

3. Conclusion

The plant *Abrus precatorious Linn* produced different pharmacological activity eg. Antimalarial, Anidiabetic, Antiinflammatory, Immunomodulator, Nephroprotectie etc. the plant also have traditional value such as aphrodisiac, remove biliousness, useful in eye diseases, cures leucoderma, itching, skin diseases and wounds. The above medicinal value of this plant is due to the presence of glycosides and alkaloids obtained from the various parts of this plant.

Reference

- [1]. Niyogi SK., Deadly crab's eye: Abrus precatorius poisoning. The New England Journal of Medicine 1969;28:51.
- [2]. Buchanan E., Grove man dies after eating rosary beans. Miami Hearld 1976; Miami FI, USA.
- [3]. Hart M. Hazards to health, Jequirity-bean poisoning, The New England Journal of Medicine 1963;268:885-86.

- [4]. Rajaram M., Janardhanan K., The chemical composition and nutritional potential of the tribal pulse *Abrus precatorius* L. Plant Foods for Human Nutrition 1992;42:4:285-90.
- [5]. Ohba H., Morowaki S. et al., Plant derived abrin A induces apoptosis in cultured leukemic cell lines by different mechanisms, Toxicology and Applied Pharmacology 2004;195:2:182-93.
- [6]. Wallis TE., Textbook of Pharmacognosy, CBS Publishers and Distributers 1998;386.
- [7]. Kapoor VK. and Handa SS., Pharmacognosy, Vallabh Prakashan 1995;104.
- [8]. William charles Evans., Trease & Evans Pharmacognosy, 15th ed., Published by Elsevier, Elsevier India Private Ltd 2002;470,186,26,186,475.
- [9]. http://medplants.blogspot.in/search/label/Abrus%20 precatorius
- [10]. Bhatia M., Siddiqui NA., Gupta SA., Precatorius (L.): An Evaluation of Traditional Herb, Indo American Journal of Pharmaceutical Research 2013;3:4:3295-3315.
- [11]. Sethi N., Nath D., Singh RK., Teratological aspects of Abrusprecatorius seeds in rats, Fitoterapia 1990;61:1:61– 63.
- [12]. Popli SP., Screening of Indian indigenous plants for antifertility activity, Progress report on project 74219(WHO), Dec. 20, 1977.
- [13]. Prakash AO. and Mathur R., Screening of Indian plants for antifertility activity, Indian Journal of Experimental Biology 1976;14:623-626.
- [14]. Krupe M., Wirth W., Nies D., Ensgraber A., Studies on the "Mitogenic" effect of hemoglutinating extracts of various plants on the human small lymphocytes in peripheral blood cultured *in-vitro*, Z Immunitatsforsch Allerg Klin Immunol 1968;135:1:19-42.
- [15]. Das PC., Sarkar AK., Thakur S., Studies on animals of a Herbo-Mineral compound for long acting contraction, Fitoterapia 1987;58:4:257-261.
- [16]. Molgaard P., Nielsen SB., Rasmussen DE., Drummond RB., Makaza N., Andreassen J., Anthelmintic acreening of Zimbabwean plants traditionally used against schistosomiasis, Journal of ethanopharmacology 2001;74:3:257-264.
- [17]. Ibrahim AM., Anthelmintic activity of some Sudanese Medicinal Plants, Phytotherapy Research 1992;6:3:155-157.

- [18]. Zia-Ul-Haque A., Qazi MH., Hamdard ME., Studies on the antifertility properties of active components isolated from the seeds of Abrusprecatorius Linn. 1. Pakistan Journal of Zoology 1983;15:2:129-139.
- [19]. Samad F., Mukhtar A., Jan ZA., Khan ZU., Effect of alcohol extract of Ratti seeds (*Abrus precatorius*) on the reproduction of female rats, Journal of Mathematical Sciences 1974;12:157.
- [20]. Bhaduri B., Ghose CR., Bose AN., Moza BK., Basu UP., Antifertility Activity of some Medicinal Plants, Indian Journal of Experimental Biology 1968;6:252,253.
- [21]. Rao MV., Antifertility effects of alcoholic seeds extract of *Abrus precatorius* Linn. in male albino rats, Acta Europaea fertilitatis 1987;18:3:217-220.
- [22]. Sinha R., Post-testicular antifertility effects of *Abrus* precatorius seed extract in albino rats, Journal of Ethnopharmacology 1990;28:2:173-181.
- [23]. Das PC., Oral contraceptive (Longacting), Patent-Brit-1445599, 1976;11.
- [24]. Agarwal SS., Ghatak N., Arora RB., Antifertility activity of the roots of *Abrus precatorius*, Pharmacological Research Communications 1970;2:159-164.
- [25]. Jadon A., Mathur R., Effects of *Abrus precatorius* Linn. seed extract on biochemical constituents of male mice, Journal of Jiwaji University 1984;9:1:100-103.
- [26]. Nwodo OFC., Botting JH., Uterotonic activity of extracts of the seeds of *Abrus precatorius*, Planta Medica 1983;47:4:230-233.
- [27]. Anon A., Barefoot Doctors's Manual, Revised Edition, Cloudburst Press of America, 2116 Western Ave., Seattle, Washington, USA. (ISBN-0-88930-012-7) Book 1977.
- [28]. Sirsi M., *In-Vitro* study of the inhibitory action of some chemotherapeutic agents on a freshly isolated strain of *Cryptococcus neoformans*, Hindustan antibiotics bulletin 1963;6:2:39-40.
- [29]. Prakash AO., Mathur R., Screening of Indian plants for antifertility activit, Indian Journal of Experimental Biology 1976;14:623-626.
- [30]. Nath D., Sethi N., Singh RK., Jain AK., Commonly used Indian abortifacient plants with special reference to their teratologic effects in rats, Journal of Ethnopharmacology 1992;36:2:147-154.
- [31]. Munsho SR., Shetye TA., Nair RK., Antifertility activity of three indigenous plant preparations, Planta Medica 1977;31:73-75.

- [32]. Prakesh AO., Gupta RB., Mathur R., Effect of oral doses of *Abrus precatorius* Linn. seeds on the oestrus cycle, body weight, uterine weight and cellular structures of uterus in albino rats, Probe 1980;19:286-292.
- [33]. Misra DS., Sharma RP., Soni BK., Toxic and haemagglutinating properties of *Abrus precatorius*, Indian Journal of Experimental Biology 1966;4:161.
- [34]. Khan AH., Gul B., Rahman MA., The interactions of the erythrocytes of various species with agglutinins of *Abrus precatorius*, Journal of Immunology 1966;96:554.
- [35]. Satyanarayana K., Sukumar K., Phytosterilants to control the cotton bug, *Dysdercus cingulatus* F, Current Science 1988;57:16:918-919.
- [36]. Krupe M., Wirth W., Nies D., Ensgraber A., Studies on the "Mitogenic" effect of hemoglutinating extracts of various plants on the human small lymphocytes in peripheral blood cultured *in-vitro*. Z Immunitatsforsch Allerg Klin Immunol 1968;135:1:19-42.
- [37]. Kusumoto IT., Kakiuchi N., Hattori M., Namba T., Sutardjo S., Shimotohno K., Screening of some Indonesian Medicinal plants for inhibitory effects on HIV-1 Protease, Shoyahugaku Zasshi 1992;46:2:190-193.
- [38]. Kusumoto IT., Shimada I., Kakiuchi N., Hattori M., Namba T., Supriyatna S., Inhibitory effect of Indonesian plant extracts on reverse transcriptase of an RNA tumour virus (1), Phytotherapy Research 1992;6:5:241-244.
- [**39**]. Nwodo OFC., Studies on *Abrus precatorius* seeds. 1. Uterotonic activity of seed oil, Journal of Ethnopharmacology 1991;31:3:391-394.
- [40]. Simpson KS., Banerjee PC., Cases of poisoning in the horse with Ratti seeds (*Abrus precatorius*) by oral administration, Indian veterinary science & animal husbandry 1932;2:59.
- [41]. Omer SA., Ibrahim FH., Khalid SA., Adam SEI., Toxicological interactions of *Abrus precatorius* and *Cassia*

senna in the diet of Lohmann broiler chicks, Veterinary and human toxicology 1992;34:4:310–313.

- [42]. Niyogi SK., Rieders F., Toxicity studies with fractions from *Abrus precatorius* seed kernels, Toxicon 1969;7:211.
- [43]. Hart M., Hazards to health, Jequiritybean poisoning, New England Journal of Medicine 1963;268:885.
- [44]. Gunsolus JM., Toxicity of Jequirity beans, Journal Amer Medical Association 1955;157:779.
- [45]. Choi YH., Hussain RA., Pezzuto JM., Kinghorn AD., Morton JF., Abrososides A-D, four novel sweet-tasting triterpene glycosides from the leaves of *Abrus precatorius*, Journal of Natural Products 1989;52:5:1118-1127.
- [46]. Canella CFC., Tokarnia CH., Dobereiner J., Experiments with plants supposedly toxic to cattle in Northeastern Brazil, with negative results.
- [47]. Apul BS., Mali JK., Poisoning of livestock by some toxic plants, Progressive Farming 1982;6:7:48.
- [49]. Jamwal KS., Anand KK., Preliminary screening of some reputed abortifacient indigenous plants, Indian Journal of Pharmaceutical Sciences 1962;24:218-220.
- [50]. Hikino H., Aota K, Takemoto T., Structure and absolute configuration of cyperotundone, Chemical and Pharmaceutical Bulletin, 1966;14:890.
- [51]. Monago CC., Alumanah EO., Antidiabetic effect of chloroform–methanol extract of *A. precatorius* seeds in alloxan diabetic rabbit 2005;9:1:85-88.
- **[52].** Ouattara K., Tuo K., Doumbia idrissa and coulibaly adama, Evaluation of the antibacterial activity of the aqueous extrct alkaloial and flavonoids from *A. precatorius* Linn. JOCPR 2012;4:11:4795-4799.
- **[53].** Rajani A., Hemamalini K., Afifa Begum, Spandana SK., Parvathalu KVLD., Gowtham, Anthelminitic activity of Ethanolic seed extract of *A. precatorius* Linn, The pharma 2013:1:11.

Source of support: Nil, Conflict of interest: None Declared

All © 2014 are reserved by International Journal of Pharmaceutical and Medicinal Research